



# Genomic prediction for tolerance to Al in a synthetic population of upland rice

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## Why Al tolerance?

In tropical South America, 85% of the soils are acidic and the production of most of staple crops is negatively impacted by acid soils





Aluminum (Al) is a metal naturally abundant in the soil, which, when pH values fall below 5.0, becomes highly phytotoxic by adopting a trivalent form  $(Al^{3+})$ 

## Why upland rice?

Upland rice (tropical japonica) is tolerant to acid soils an important component of rice-pasture cropping systems.

Rice-pasture cropping system, a productive option for the savannas

Since 1992 the Cirad-CIAT rice breeding program has worked on improving upland rice for the acid savannas of Colombia and LAC



Soils improvement as a strategy to enhance productivity in agricultural systems of the acid savannas





## Recombination among best progenies to improve the population value

### Why Recurrent Selection on synthetic populations?

To combine many required trait of interest.

Synthetic populations can thus be improved with a particular focus for adaptation to the savanna soils.

The recurrent selection method of plant breeding carries the advantage to target the improvement of quantitative traits such as those usually controlling adaptative characters.



Mean value increased without compromising the genetic variability



### Why Genomic Prediction aided RS?

Accelerate the development of elite material

Operate more precise selection

Save cost in the breeding program

Increase genetic gain

 $\Delta G_{Yr} = \frac{i_{NS} \cdot r_{g\hat{g}} \cdot \sigma_{A}^{2}}{I}$ 



## Our approach

Families derived from a synthetic population which is the base of the upland rice breeding program

Tropical japonica germplasm derived from the recombination of elites upland rice from Asia, Africa and Latin America and three decades of recombination



PCT27 generated from the mixture of for population (Baertschi et al., 2021) Genotyping performed at  $S_0$  generation 334 advanced generation families ( $S_{0:4}$ ) randomly drawn from the PCT27

### **Experimental set up**

- 334 S<sub>0:4</sub> lines (grey colour)
- + 25% repetitions = 84 S<sub>0:4</sub> lines (yellow colour)
- + 50  $S_{0:2}$  lines (for cross year comparisons & GxE-Genomic Prediction)
- + 6 checks repeated 6 times (red colour)
- → 504 plots (6 ranges with 6 blocks of 14 plots)

4 3m long row 25 cm between rows Sowing density: 1 g/m



### **Data Analysis**

BLUEs estimation using mixed linear model

Calibration model (BGGE R-CRAN) by Gustavo de los Campos & Paulino Perez-Rodriguez)

pTst <- 0.3 folds <- 100 nlter <- 70000; burnIn <- 2000

#### ➔ GEBVs



Predictive ability (PA) = r (GEBVs, BLUEs) from fold 1 to 100

### BGGE – the Bayesian linear mixed models (Granato et al., 2018)

SM : single-environment, main genotypic effect model -- fits the data for a single environment, and only one kernel is produced

Model	Main genetic effect of line across environments	Genotype × environment interaction (G×E)				
MM	$Z_{u}KZ'_{u}$					
MDs	$Z_{u}KZ'_{u}$	$(Z_{u}KZ'_{u})^{\circ}Z_{E}Z'_{E}$				
MDe	$Z_u K Z'_u$	$\begin{bmatrix} 0 & \cdots & 0 & \cdots & 0 \\ \vdots & \ddots & \vdots & \ddots & \vdots \\ 0 & \cdots & K_j & \cdots & 0 \\ \vdots & \ddots & \vdots & \ddots & \vdots \\ 0 & \cdots & 0 & \cdots & 0 \end{bmatrix}$				
		for each environment $j (j = 1,, m)$				

MM : multi-environment, main genotypic effect model -- considers the main random genetic effects across environments.

MDs : multi-environment, single variance GxE deviation mode -extension of MM by adding the random interaction effect of environments with genotype information.

MDe : multi-environment, environment-specific variance GxE deviation model -- separates the genetic effects into the main genetic effects and the specific genetic effects (for each environment).

Zu is a known incidence matrix that relates the genotypes to the observations in the environments

## Trait distribution in the whole set







# Effect of treatment on grain yield

Greater general performance of indica rice under any treatment

Only within the PCT27 grain yield significantly greater under Al than Cal

→ Upland rice families have lower yield than the indica rice but tolerance to aluminum seems to exist in this set of germplasm



### High trait correlation and variability in AITol index





Potential to exploit AlTol among the families

### GP on the 334 S<sub>0:4</sub> families for trait in Al condition



Predictive ability as the Pearson correlation between predicted values (GEBVs) and the estimated values (BLUEs)

Large variation in PA according to the traits

MDe

MDs MM

SM

Considering GxE has different impact on the PA

Generally better PA when using the data from both sites to calibrate the model

### **High PA for FL considering** multisite models

SM



	FL	Df	Sum Sg	Mean Sq	F value	Pr(>F)	
	DESIGN	383	18837.1	49.18	18.7591	0.051905	
	SITE	1	652.9	652.91	249.0299	0.003992	**
	DESIGN:SITE	383	4688.9	12.24	4.6695	0.192681	
	SITE:R	10	476.2	47.62	18.1612	0.053289	
el MDe	SITE:C	156	1497.1	9.6	3.6603	0.238695	
MDs	Residuals	2	5.2	2.62			
MM							

Low PA with the single-environment, best when accounting for the GxE and genotypic as main effect or with single GxE variance effect





High PA regardless of the model probably due to the low GxE contribution and high phenotypic correlation between sites

#### Model

MDs

MM

SM

## Good PA for YLD with main genetic effect



	Groups	Variance	Std.Dev.	$\left. \sigma_{ge}^2 \right/ \!$
FL	DESIGN:SITE	1.9938	1.412	0.13
	DESIGN	15.0419	3.8784	
	Residual	4.0519	2.0129	
PH	DESIGN:SITE	2.51E-12	1.58E-06	4E-14
	DESIGN	55.97	7.48	
	Residual	32.16	5.67	
YLD	DESIGN:SITE	0	0	0.00
	DESIGN	212260	460.72	
	Residual	149032	386.05	
ZN	DESIGN:SITE	1.3394	1.1573	1.22
	DESIGN	1.0947	1.0463	
	Residual	4.2442	2.0602	

Greater PA with multi-environment, main genotypic effect model probably because variance explained by GxE remains low compared to the genetic variance



Good PA of ZN with



High PA for ZN with GxE and main genotypic effect explained as high  ${\rm H}^2$ 

### Discussion

Although lower in productivity than the indica rice the potential of japonica is important to consider for acid soils.

Japonica rice can have a function for degraded soil restauration, crop rotation with pasture/leguminous.

Japonica rice can bring interesting gene to high yielders indica rice.

Good perspective to increase genetic gain in the breeding program through genomic prediction aided recurrent selection.



#### **Next step -- Genetics**

The GWAS combined with the genomic prediction could potential improve the fixation of certain key genes

Need of more markers (GBS pending) to detect more closely the responsible genetic control



Finer evaluation of GP and estimate on intra family variance to further the optimization of the breeding scheme to adaptation to acid soils

## **Next steps -- Connections**

The 50 elite lines evaluated for drought and Al tolerance are derived from populations which conformed PCT27. Validation of genetic control in the 50 lines can be done on this larger population

- → direct application of MAS with targeted regions / combination of GP and single marker MAS to combine several traits of interest
- $\rightarrow$  discovery of new mechanisms (under more quantitative control)

General poor performance of the tropical japonicas for lack of resistance to RHBV

- → should be taking advantage of the work on other P6 for general improvement of upland rice Population available for further phenotyping of quantitative traits and that is of direct use for breeding
- ightarrow consider a subset for phenotyping climate resilient traits



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